

Coexpression of ABCB1 and ABCG2 as prognostic marker for treatment response in acute myeloid leukemia patients

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Received: May 30, 2022

Accepted: Sep 01, 2022

Published: Sep 09, 2022

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Keywords: ABCB1;
ABCG2;
Acute Myeloid Leukemia;
Flowcytometry.

Abstract

Treatment resistance is nowadays the main challenge in the field of AML. ABC transporters are considered one of the main mechanisms of treatment resistance in AML. Moreover, its prognostic impact on short and long-term remission rate remains of great concern. The development of targeting therapy is an urgent need for improving remission rate and prolongs survival in AML patients. This study aimed at assessing the relationship between co-expression of ABCB1 and ABCG2 transporters and their relation with the response to induction therapy as well as their prognostic significance in newly diagnosed AML patients.

This study included forty newly diagnosed AML patients and thirty healthy subjects of matched age and sex as a control group. Complete blood count, and Flowcytometric measurement of ABCB1 (CD 243) and ABCG2 (CD 338) were done for all study participants while Bone marrow aspiration (BMA) was carried out for AML patients only. Patients were reevaluated after 28 days from receiving induction treatment in the form of (3+7) protocol for remission achievement. A statistically significant correlation between co-expression of ABCB1 and ABCG2 and treatment response was observed. This denoted that ABCB1 and ABCG2 co-expression can be used as a prognostic marker for remission achievement in AML patients. However, further studies are still needed to confirm this finding in Egyptian population.

Citation: Mahmoud A, Elghandour A, Elhadidi A, Abdelrahman A, Eldafrawi M. Coexpression of ABCB1 and ABCG2 as prognostic marker for treatment response in acute myeloid leukemia patients. BAOJ Cancer Res The. 2022; 6(1): 1003.

Introduction

Acute Myeloid Leukemia (AML) is the most common type of acute leukemia, and despite availability of chemotherapeutic drugs, mortality rates remain highly variable, ranging from 90 to 10% [1]. One of main characters of AML is having distinct cytogenetic and molecular subgroups. Despite trials trying to improve short- and long-term remission rate, fate of AML patients remains doubtful, resistance and relapse rates remains high. Many questions have been addressed to solve the problem of drug resistance [2].

Treatment resistance remains an obstacle and a challenge to hematologist. Many mechanisms of resistance have been described in AML including functional mechanism in leukemic blasts or due to host or acquired factors [3].

One of the main mechanism of resistance is ATP membrane transporter responsible for chemotherapeutic drug efflux as (ABCB1, ABCG2,..... etc). Overexpression of these transporters has a poor prognostic impact on remission rates in acute myeloid leukemia. Coexpression of multiple ABC transporters results in a worse prognosis [4].

P-glycoprotein is one of the most studied ABC transporters which has a role in drug resistance. It is encoded by the multidrug resistance gene 1 [5]. Other ABC-transporters with clinical impact on drug resistance in adult AML are the breast cancer resistance protein (BCRP), encoded by adenosine triphosphate binding cassette transporter G2 (ABCG2) [6].

The increased expression of ABC transporters causes efflux of chemotherapeutic drugs out of leukemic cells and reducing the effectiveness of these drugs resulting in increasing of the resistance and relapse rate in these patients [7-9]. Approximately 50% of AML patients express ABC transporters, with increased level of expression in elderly and relapsed patients [3]. Indeed, more than one ABC transporters is expressed in leukemic blast of patients especially relapsed patients [10].

Fortunately, clinical trials paid a great attention to overcome problem of drug resistance through developing ABC transporters inhibitors. Indeed, with the appearance of these new data about possibility of developing target therapy, we propose to reassess the role of these transporters in multidrug resistance [11-13].

The aim of this study was to investigate the expression levels of ABCB1 and ABCG2 and their effect on the achievement of hematological remission after induction therapy as well as their prognostic value in newly diagnosed AML patients.

Patients and methods

The present study was conducted on 40 newly diagnosed AML patients, received 3+7 protocol as induction therapy. Assessment of hematologic remission was carried out after 28 days. Flowcytometric measurement of CD243 and CD338 was done at diagnosis. Patients were recruited from the Hematology outpatient clinic of Alexandria Main University Hospital during the period from May 2016 to December 2017.

Thirty healthy subjects of matched age and sex were included in the study as a control group. Patients with hepatic or renal failure, concomitant chronic illness and pregnant females were excluded from the study. Written consent was taken from all study participants. This study was approved by the Alexandria Faculty of Medicine Ethics Committee.

All subjects included in the study were subjected to the following:

1. Complete history taking.
2. Thorough clinical examination.
3. Laboratory investigation including:
 1. Routine Lab Investigation:
 - Complete blood count.
 - ALT, AST, S.urea, S.creatinine.
4. Bone marrow aspiration and examination.
5. Immunophenotyping by flowcytometry.

Immunophenotyping of the leukemic blast cells was performed on bone marrow samples using BD FACSCalibur flow cytometry analyzer equipped with BD Cell Quest Pro Software.

The following McAbs were used in combination.

The following panel applied for all cases (CD2, CD7, CD10, CD14, CD19, CD13, CD33, HLD-DR, CD34, CD45, CD 11c).

- a) Secondary panel: CD11b, CD64, cyt MPO, cyt CD 22, cyt CD 3, CD 117, CD 41, CD 61, CD 235a.
- b) Analysis of CD 243 (ABCB1) CD 338 (ABCG2) expression on leukemic blasts of AML by flowcytometry.

ABCB1 (CD 243) PE, Clone: 41C2, Catalog number: 12-2439-42Lot number: 13, Product of Thermo-Fischer, scientific, San Diego.

ABCG3 (CD 338), Clone: 5D3, Catalog number: 13-8888-82, Lot number: 114531, Product of Thermo-Fischer, scientific, San Diego. Labelled by APC.

Results

Regarding demographic data, there was no statistically significant difference between patients and controls as shown in table 1.

Table 1: Comparison of demographic data between patients and controls.

Age (years)	Groups				t-Test p
	Patients	Control			
Range	25.0-52.0	23.0-50.0			2.196
Mean	38.9	35.8			0.143
S.D.	8.3	8.6			N.S.
Sex	No.	%	No.	%	0.152
Male	29	72.5	23	62.5	
Female	11	27.5	7	37.5	

P is significant if ≤ 0.05

When testing conventional cytogenetics in relation to CD 338 expression and CD 243, it was noted that there is statistically significant difference between the level of both CD expression and karyotyping ($p= 0.037$, $p= 0.001$) respectively as shown in table 2 and 3.

Table 2: Relation between conventional cytogenetics in relation to CD338 expression.

Cytogenetics		CD338		Total
		-ve	+ve	
Complex	No.	5	3	8
	%	8.5%	27.3%	11.4%
Hyperdiploidy	No.	0	1	1
	%	0.0%	9.1%	1.4%
Inversion 16	No.	1	1	2
	%	1.7%	9.1%	2.9%
Normal	No.	19	1	20
	%	32.2%	9.1%	28.6%
t (8;21)	No.	4	0	4
	%	6.8%	0.0%	5.7%
failure to obtain	No.	5	0	5
	%	8.5%	0.0%	7.1%
X ²		12.554		
p		0.037*		

Table 3: Relation between conventional cytogenetics in relation to CD243 expression.

Cytogenetics		CD243		Total
		-ve	+ve	
Complex	No.	7	1	8
	%	10.6%	25.0%	11.4%
Hyperdiploidy	No.	0	1	1
	%	0.0%	25.0%	1.4%
Inversion 16	No.	2	0	2
	%	3.0%	0.0%	2.9%
Normal	No.	19	1	20
	%	28.8%	25.0%	28.6%
t8:21	No.	3	1	4
	%	4.5%	25.0%	5.7%
Failure to obtain	No.	5	0	5
	%	7.6%	0.0%	7.1%
X ²		12.554		
p		0.037*		

Regarding Treatment, According to bone marrow aspirate results, the following table (4) shows response to standard treatment protocol (3+7) among our patients.

Table 4: Response to the planned treatment among AML cases.

	Number	Percent
Remission (CR)	26	65.0
Partial remission (PR)	14	35.0
Total	40	100.0

After induction treatment, CD 243 expression in patients who achieved complete response was negative in 100% and positive in zero %, regarding patients with partial remission was negative in 71.4% and positive in 28.6% and this was statistically significant (p < 0.05). As regards CD 338 expression in patients who

achieved complete response after induction chemotherapy was negative in 96.2% and positive in 3.8%, regarding patients with partial remission was negative in 64.3% and positive in 35.7% and this was statistically significant (p<0.05) as seen in table 5.

Table 5: Relation between the CD 338 and CD 243 expression and response to treatment.

		Response		X ² P	
		Complete response	Partial response		
CD338	-ve	No.	25	9	7.248 0.014*
		%	96.2%	64.3%	
	+ve	No.	1	5	
		%	3.8%	35.7%	
CD243	-ve	No.	26	10	4.13 0.035*
		%	100.0%	71.4 %	
	+ve	No.	0	4	
		%	0.0%	28.6 %	

Figures (1-3) show different patterns of expression of CD 243 and CD 338 in three of our patients.

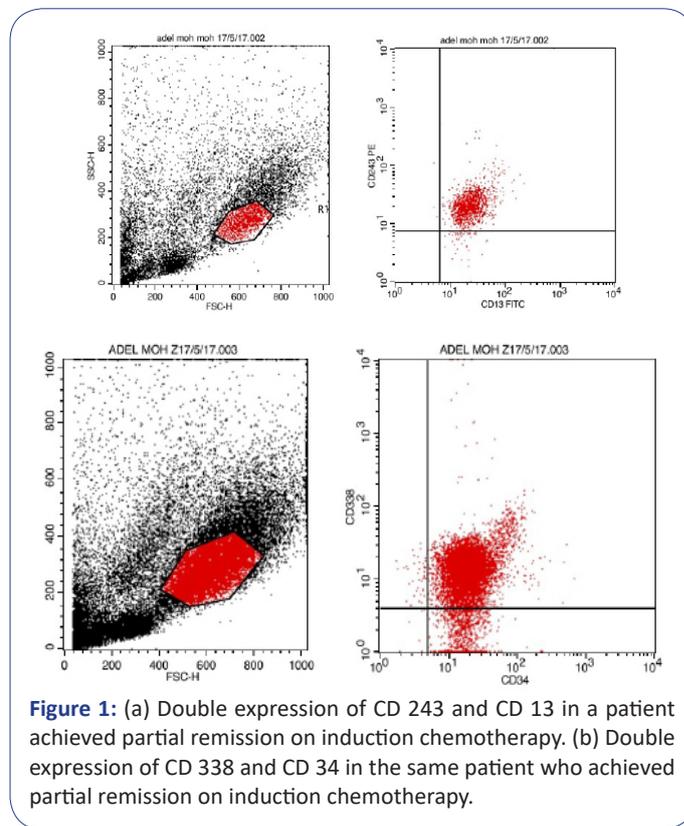


Figure 1: (a) Double expression of CD 243 and CD 13 in a patient achieved partial remission on induction chemotherapy. (b) Double expression of CD 338 and CD 34 in the same patient who achieved partial remission on induction chemotherapy.

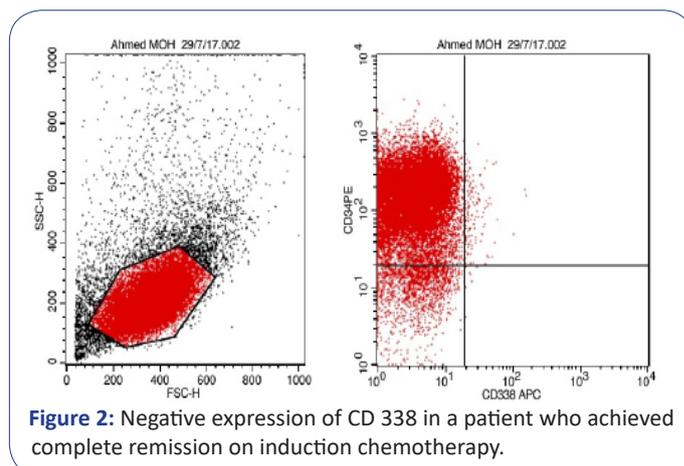


Figure 2: Negative expression of CD 338 in a patient who achieved complete remission on induction chemotherapy.

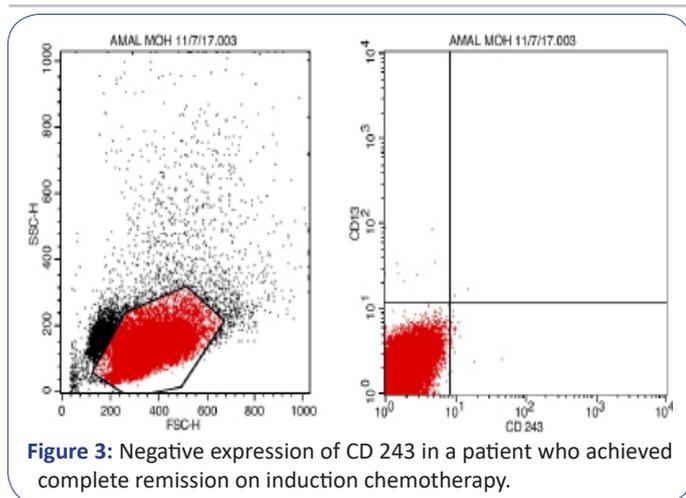


Figure 3: Negative expression of CD 243 in a patient who achieved complete remission on induction chemotherapy.

Discussion

Coexpression of multiple ABC transporters carries the worse prognosis in adulthood AML patients and contribute to treatment resistance which remains a challenge in those patients [4]. MDR is a multifactorial process including the following mechanisms as i) altered membrane transport. ii) altered target enzymes. iii) altered drug activation or degradation. iv) enhanced DNA repair. v) failure to undergo apoptosis. vi) abnormal autophagy [14].

Effectiveness of chemotherapeutic drugs depends on transport mechanism (drug efflux) through ATP transporters which remains one of the main mechanisms contributing to treatment resistance [15].

The ABC superfamily consists of 49 ABC genes identified in the human genome. Currently at least 15 ABC transporters have been implicated to confer resistance to clinically active drugs, notably P-glycoprotein (P-gp, ABCB1) and breast cancer resistance protein (BCRP, ABCG2) [16].

Resistance conferred by ABC transporters is not only to the classic chemotherapeutic agents but also against the new targeted therapy. Also many researchers discussed interactions between ABC transporters and target therapy [17-20].

Coexpression of Multiple ABC-Transporters is strongly associated with inferior outcomes in AML. Researchers' main concern nowadays is inhibiting these transporters which will improve the short term response and decrease relapse rate [21].

To date, many ABCB1 inhibitors have been developed, such as cyclosporine A. But astonishing, inhibiting ABCB1 transporter only was associated with inferior results and did not overcome MDR. THIS raised into mind the idea about co-expression of multiple ABC transporters has more great impact on treatment resistance [22-25].

We designed our study depending on this point of view, we concluded that coexpression of ABCB1 and ABCG 2 rather than one transporter is associated with poor prognosis in newly diagnosed AML patients after induction chemotherapy.

Declarations

Ethics approval and consent to participate: All patients provided written informed consent prior to their inclusion within the study.

Consent for publication: All patients provided written informed consent for the publication of their data.

Competing interests: The authors declare that they have no competing interests.

Author contributions: This study and research were designed by AM, AG, AE, AA, ME. The experiments were conducted and analyzed by AM and AE, flow cytometry by AE. Reagents were provided by AM. The manuscript was written by AM, AG, AE.

Financial Disclosure Statement: no financial conflict of interest to disclose.

Conflicts of interest: The authors declare that they have no competing interests.

Acknowledgments: I thank my professor,mentor and godfather Prof Ashraf Elghandour who is holding my hand and lighting my way. Team work members thank Prof Mohamed Eldafrawi for his great share and assistance in designing this study and follow up work closely, may God bless his soul. We thank Clinical Pathology department for great support in the study and providing the required equipments. We thank the technicians and nurses in Alexandria Main University Hospitals Hematology department for technical support.

References

1. Kayser S, Zucknick M, Döhner K, Krauter J, Köhne CH, et al. Monosomal karyotype in adult acute myeloid leukemia: prognostic impact and outcome after different treatment strategies. *Blood*. 2012; 119: 551-558.
2. Yeung CCS, Radich J. Predicting Chemotherapy Resistance in AML. *Curr Hematol Malig Rep*. 2017; 12: 530-536.
3. Shaffer BC, Gillet JP, Patel C, Baer MR, Bates SE, et al. Drug resistance: still a daunting challenge to the successful treatment of AML. *Drug Resist Updat*. 2012; 15: 62-69.
4. Liu B, Li LJ, Gong X, Zhang W, Zhang H, et al. Co-expression of ATP binding cassette transporters is associated with poor prognosis in acute myeloid leukemia. *Oncol Lett*. 2018; 15: 6671-6677.
5. Steinbach D, Legrand O. ABC transporters and drug resistance in leukemia: was P-gp nothing but the first head of the Hydra? *Leukemia*. 2007; 21: 1172-1176.
6. Damiani D, Tiribelli M, Calistri E, Geromin A, Chiarvesio A, et al. The prognostic value of P-glycoprotein (ABCB) and breast cancer resistance protein (ABCG2) in adults with de novo acute myeloid leukemia with normal karyotype. *Haematologica*. 2006; 91: 825-828.
7. Marquez B, Van Bambeke F. ABC multidrug transporters: target for modulation of drug pharmacokinetics and drug-drug interactions. *Curr Drug Targets*. 2011 12(5):600-620.
8. Hourigan CS, Karp JE. Personalized therapy for acute myeloid leukemia. *Cancer Discov*. 2013; 3: 1336-1338.
9. Anreddy N, Patel A, Zhang YK, Wang YJ, Shukla S, et al. A-803467, a tetrodotoxin-resistant sodium channel blocker, modulates ABCG2-mediated MDR in vitro and in vivo. *Oncotarget*. 2015; 6: 39276-39291.
10. Walter RB, Raden BW, Cronk MR, Bernstein ID, Appelbaum FR, Banker DE. The peripheral benzodiazepine receptor ligand PK11195 overcomes different resistance mechanisms to sensitize AML cells to gemtuzumab ozogamicin. *Blood*. 2004; 103: 4276-4284.
11. Castro J, Ribó M, Puig T, Colomer R, Vilanova M, Benito A. A cytotoxic ribonuclease reduces the expression level of P-glycoprotein in multidrug-resistant cell lines. *Invest New Drugs*. 2012; 30: 880-888.

12. Lainey E, Wolfrohm A, Marie N, Enot D, Scoazec M, et al. Azacytidine and erlotinib exert synergistic effects against acute myeloid leukemia. *Oncogene*. 2013; 32: 4331-4342.
13. Robey RW, Pluchino KM, Hall MD, Fojo AT, Bates SE, Gottesman MM. Revisiting the role of ABC transporters in multidrug-resistant cancer. *Nat Rev Cancer*. 2018; 18: 452-464.
14. Bugde P, Biswas R, Merien F, Lu J, Liu DX, et al. The therapeutic potential of targeting ABC transporters to combat multi-drug resistance. *Expert Opin Ther Targets*. 2017; 21: 511-530.
15. Fukuda Y, Schuetz JD. ABC transporters and their role in nucleoside and nucleotide drug resistance. *Biochem Pharmacol*. 2012; 83: 1073-1083.
16. Li W, Zhang H, Assaraf YG, Zhao K, Xu X, et al. Overcoming ABC transporter-mediated multidrug resistance: Molecular mechanisms and novel therapeutic drug strategies. *Drug Resist Updat*. 2016; 27: 14-29.
17. Bhullar J, Natarajan K, Shukla S, Mathias TJ, Sadowska M, et al. The FLT3 inhibitor quizartinib inhibits ABCG2 at pharmacologically relevant concentrations, with implications for both chemosensitization and adverse drug interactions. *PLoS One*. 2013; 8: e71266.
18. Hupfeld T, Chapuy B, Schrader V, Beutler M, Veltkamp C, et al. Tyrosinekinase inhibition facilitates cooperation of transcription factor SALL4 and ABC transporter A3 towards intrinsic CML cell drug resistance. *Br J Haematol*. 2013; 161: 204-213.
19. Grundy M, Seedhouse C, Russell NH, Pallis M. P-glycoprotein and breast cancer resistance protein in acute myeloid leukaemia cells treated with the aurora-B kinase inhibitor barasertib-hQPA. *BMC Cancer*. 2011; 11: 254.
20. Chapuy B, Panse M, Radunski U, Koch R, Wenzel D, et al. ABC transporter A3 facilitates lysosomal sequestration of imatinib and modulates susceptibility of chronic myeloid leukemia cell lines to this drug. *Haematologica*. 2009; 94: 1528-1536.
21. Jaramillo AC, Saig FA, Cloos J, Jansen G, Peters GJ. How to overcome ATP-binding cassette drug efflux transporter-mediated drug resistance? *Cancer Drug Resist*. 2018; 1: 6-29.
22. Marzac C, Garrido E, Tang R, Fava F, Hirsch P, et al. ATP Binding Cassette transporters associated with chemoresistance: transcriptional profiling in extreme cohorts and their prognostic impact in a cohort of 281 acute myeloid leukemia patients. *Haematologica*. 2011; 96: 1293-1301.
23. Hauswald S, Duque-Afonso J, Wagner MM, Schertl FM, Lübbert M, et al. Histone deacetylase inhibitors induce a very broad, pleiotropic anticancer drug resistance phenotype in acute myeloid leukemia cells by modulation of multiple ABC transporter genes. *Clin Cancer Res*. 2009; 15: 3705-3715.
24. Bartholomae S, Gruhn B, Debatin KM, Zimmermann M, Creutzig U, et al. Coexpression of Multiple ABC-Transporters is Strongly Associated with Treatment Response in Childhood Acute Myeloid Leukemia. *Pediatr Blood Cancer*. 2016; 63: 242-247.
25. Liu Y, Cui P, Chen J, Li W. Isolation and phenotypic characterization of side population cells in oral squamous cell carcinoma. *Mol Med Rep*. 2015; 11: 3642-3646.