

Mini Review**Plant Secondary Metabolites: Natural Antibacterial Agents**

Anil Kumar*

Abstract

Secondary metabolites are produced by the plants and these include alkaloids, phenolics, flavonoids, etc. Unlike primary metabolites which are directly involved in plant growth, development, and reproduction, these secondary metabolites are considered to be defense tools for the plant. In the last decade, much research has been carried out on the antibacterial activity of these plant secondary metabolites. This article discusses several secondary metabolites viz. alkaloids, terpenoids, flavonoids, and phenolics for their antibacterial activities.

Introduction

The metabolites play important role in the physiology and biochemistry of the organisms. Any living organism can't survive without metabolites. The metabolites have role in maintaining the structure, and/ or may work as a source of energy[1]. Besides, certain metabolites also act as signalling molecules[2], effectors for various enzymes[3], co-factor for the enzyme[4, 5], and defense molecules[6]. The metabolites are of two different categories, namely primary and secondary metabolites, depending on their role in the organism. If the metabolite is directly involved in the growth, development, reproduction, etc. of the organism, it is called primary metabolite[7]. When a metabolite is not directly involved in growth, development, reproduction, etc., then it is called secondary metabolite[7]. The secondary metabolites are not the obligate requirement for the survival of an organism. It has been shown that secondary metabolites are involved in long-term impairment of the organism's survivability[8].

Although other organisms also synthesize secondary metabolites, these are mainly synthesized by the plants and these have an important role in plant defense against many pathogens[9]. In recent years, this property of secondary metabolites is being exploited as natural medicines against many diseases[10]. The secondary metabolites are found in various amounts in different parts of the plants. The amount depends upon the stage of growth and under different stress conditions like presence of invasive microorganisms, herbivores, etc.[11].

Since last few decades, the side effect of synthetic medicines and resistance being developed in bacteria against many antibiotics has attracted the attention of several researchers towards this field.

The secondary metabolites are of different types such as alkaloids, phenolics, flavonoids, etc. These secondary metabolites have various roles like insect attractants for help in pollination, defense against bacteria, insects, and predators. In addition to the medicinal role, these secondary metabolites have been exploited for flavours, fragrances in food, and cosmetics industries [12-16].

Medicinal plant is a plant when it is as such or some constituent of it is exploited as a medicine for certain disease(s). Nowadays, many plants have been identified as medicinal plants due to the presence of secondary metabolites in them which act as natural antimicrobial agents. The important secondary metabolites which have been identified as natural antimicrobial agents are the following:

Alkaloids

Alkaloids are nitrogenous organic compounds. The word 'Alkaloid' was coined from alkali as mostly these compounds are alkaline in nature. However, some alkaloids have been reported having neutral or acidic property.

Alkaloids have carbon, hydrogen, and nitrogen in their structure. Besides, some of these also have oxygen, sulfur, chlorine, bromine, and phosphorus. Alkaloids are produced by the plants as their defense mechanism[19]. However, few bacteria and fungi have also been reported to produce alkaloids[20]. Alkaloids have been categorized as pharmaceuticals since various alkaloids exhibit activity against many diseases. The most popular alkaloid, quinine, has anti-malarial activity[21]. It has activity against

***Corresponding Author:** School of Biotechnology, Devi Ahilya University, Khandwa Rd, Indore-452001, India, E-mail: ak_sbt@yahoo.com

Sub Date: March 14th, 2018, **Acc Date:** March 31st, 2018, **Pub Date:** March 31st, 2018.

Citation: Anil Kumar (2018) Plant Secondary Metabolites: Natural Antibacterial Agents. BAOJ Biotech 4: 029.

Copyright: © 2018 Anil Kumar. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Plasmodium falciparum, a microbe responsible for causing malaria in humans. This is transmitted in humans through mosquitoes. Quinine was first isolated in 1820 from the bark of a cinchona tree. Bark extracts are used to treat malaria since centuries. It is on the WHO Model List of Essential Medicines as most important medicine required in a primary health centre. The *Aspidosperma olivaceum* is being used as a medicine against fevers in many parts of the world and its medicinally active ingredients have been reported to be indole alkaloids[22]. The bark and leaves of the plant have also been shown to exhibit antimicrobial activity against *Plasmodium falciparum*[22].

Vincamine is a monoterpene indole alkaloid which has been isolated from the leaves of *Vinca minor*. This plant has been reported to be enriched in vincamine which constitutes about 25-65% of the indole alkaloids. Vincamine is a peripheral vasodilator that increases blood flow to the brain[23].

Chelerythrine, a benzophenanthridine alkaloid, has an antibacterial activity against *Staphylococcus aureus* and many other human pathogens. It has been reported to be present in plants viz. *Chelidonium majus*. It is a potent, selective, and cell-permeable protein kinase C inhibitor *in vitro*. It has also been reported to be efficacious antagonist of G-protein-coupled CB1 receptors[24].

The *Catharanthus roseus* (also known as *Vinca rosea*), a plant belonging to family, Apocynaceae is known for its alkaloids, Vinblastine and vincristine, which have been reported to exhibit anti-cancerous activity[25].

Terpenoids

Azadirachtin, a terpenoid, is a secondary metabolite present in neem (*Azadirachta indica*) seeds. Earlier, it was shown to have inhibitory activity towards the desert locust (*Schistocerca gregaria*). Later, it has been reported to affect nearly 200 species of insect, by acting mainly as an anti-feedant and growth disruptor, and as such it possesses considerable toxicity towards insects. Due to its biodegradable property, Azadirachtin has been considered to be a very good insecticidal. It is degraded within 100 hours when exposed to light and water, and showed very low toxicity towards mammals[26].

Artemisia annua L., a medicinal herb has also been reported to produce secondary metabolites which exhibit antimicrobial activity. This plant has artemisinin, a terpenoid, which is the active ingredient responsible for antimicrobial activity. The secondary metabolites present in this plant are reported to have inhibitory activity against Gram-positive and Gram-negative bacteria. Kim et al.[27] reported anti-inflammatory, antioxidant, and antimicrobial properties of artemisinin derived from water, methanol, ethanol, or acetone extracts of *Artemisia annua* L. They found antimicrobial activity of the extracts against the periodontopathic microorganisms, *Aggregatibacter actinomycetemcomitans*, *Fusobacterium nucleatum* subsp. *animalis*, *Fusobacterium nucleatum* subsp. *polymorphum*, and *Prevotella*

intermedia. They recommended the use of *Artemisia annua* L. for dental diseases since these extracts also exhibited the anti-inflammatory and antioxidant activities also.

Flavonoids

Antimicrobial activity of plant flavonoids is known since long time. Mitscher et al.[28] reported isolation of number of isoflavonoids and related compounds from *Glycyrrhiza glabra* L. var. typical and identified them as glabridin, glabrol, glabrene, 3-hydroxyglabrol, 4'-O-methylglabridin, 3'-methoxyglabridin, formononetin, phaseollinisoflavan, hispaglabridin A, hispaglabridin B, salicylic acid, and O-acetyl salicylic acid. They showed that out of these, hispaglabridin A, hispaglabridin B, 4'-O-methylglabridin, glabridin, glabrol, and 3-hydroxyglabrol possessed significant antimicrobial activity *in vitro*. They also reported that hispaglabridin A, hispaglabridin B, 3'-methoxyglabridin, 4'-O-methylglabridin, 3-hydroxyglabrol, phaseollinisoflavan, salicylic acid, and O acetyl salicylic acid are newly found in *Glycyrrhiza* sp.; and hispaglabridin A, hispaglabridin B, 3'-methoxyglabridin, 4'-O-methylglabridin, and 3-hydroxyglabrol are new to the literature.

Dastidar et al.[29] studied antibacterial potential of isoflavones. They screened isoflavonoid compounds 'YS11-YS21' for possible antimicrobial property against 12 known Gram-positive and Gram-negative sensitive bacteria. They found that YS11 and YS16 failed to show antimicrobial activity whereas YS12, 13, 14, 15, 17, 18, and 20 had moderate antimicrobial action. However, compounds YS19 and YS21 showed pronounced antimicrobial property. The isoflavones YS19 and YS21 were then tested *in vitro* against 214 strains of bacteria from one Gram-positive and six Gram-negative genera. They determined the minimum inhibitory concentration (MIC) of YS19 and YS21 by agar dilution method and it ranged from 25 to 200 mg/l in most strains. At concentrations of 30 and 60 µg/mouse, these compounds offered significant protection to mice challenged with 50 median lethal dose (MLD) of a virulent strain of *Salmonella typhimurium*.

Mukne et al.[30] reported that flavonoids and related compounds possessed potent antimicrobial activities. Most of the flavonoids are considered as constitutive antimicrobial substances termed as "Phytoanticipins," especially those belonging to prenylated flavonoids and isoflavones. They highlighted the structural prerequisites for isoflavones as antibacterial agents and drew Structure-activity relationship (SAR) conclusions by comparing the reported minimum inhibitory concentration values for the various isoflavones against *Staphylococcus aureus* and methicillin-resistant *Staphylococcus aureus* (MRSA). They also formulated a significant correlation between the presence of certain functional groups viz. prenyl and phenolic hydroxyl at particular positions and antibacterial activity of the compounds. They postulated these trends with a view of assisting better drug designing of future next-generation anti-infectives,

particularly against the bothersome multidrug-resistant microbes. They showed that SAR of these isoflavones also proved to be a basis to explore the mechanism of antibacterial action. They hoped that their study would prove extremely useful to synthesize antibacterial isoflavones in future, which would eventually be beneficial for optimizing the lead molecule for the antibacterial action.

Praveen and Ghalib [31] showed that methanolic extract of the leaves of *Xylosma longifolium* has a new flavonoid compound, Kaempferol-3- β -xylopyranoside-4'- α -rhamnoside, along with Kaempferol, Quercetin, Kaempferol-3-rhamnoside, and Quercetin-3-rhamnoside. They checked the *in vitro* antimicrobial activity of the aqueous and methanolic extract against *Escherichia coli*, *Staphylococcus aureus*, *Salmonella typhimurium*, and *Bacillus subtilis*. They tested *Candida albicans*, *Fusarium oxysporum*, *Penicillium notatum*, *Aspergillus niger*, and *Trichoderma viridae* for the *in vitro* antifungal activity. They found that these extracts had antimicrobial activity against Gram-positive bacteria and fungal strains. The aqueous extract exhibited high antimicrobial activity against *Staphylococcus aureus* and *Candida albicans*, and moderate activity against *Bacillus subtilis* and *Trichoderma viridae*. The methanolic extract exhibited antimicrobial activity against *Staphylococcus aureus* and *Candida albicans*, and moderate activity against *Bacillus subtilis*, *Salmonella typhimurium*, and *Aspergillus brassicola*.

Phenolics

Phenolics are the well-known antibacterial agents. These are synthesized by the plants and are considered to be as secondary metabolites. Phenolics protect the plants from pathogenic bacteria. It has been shown that the growth of *Xylella fastidiosa*, a pathogenic bacteria which infects many crops, gets inhibited by phenolic acid as shown using low minimum inhibitory concentrations [32]. Maddox et al. [32] also showed that phenolics with different structural features viz. catechol, caffeic acid, and resveratrol exhibited strong antibacterial activity against different strains of *Xylella fastidiosa* isolated from grape and almond. Papuc et al. [33] reviewed plant polyphenolics as antibacterial agents. The phenolics are used as preservative for increasing the shelf life of meat and meat products [34, 35]. It is shown that polyphenols interact with the cell wall components of the bacteria and bacterial cell membrane control biofilm formation. Besides, phenolics are known to inhibit microbial enzymes. They also influence the regulation of protein biosynthesis. The phenolics may also deprive the bacterial cell enzymes of substrates and metal ions.

Ouerghemmi et al. [36] isolated phenolics from the leaves and flowers of Tunisian *Ruta chalepensis* L and identified vanillic acid and coumarin mainly in the extract. They extracted from cultivated as well as wild grown Tunisian *Ruta chalepensis*, and found no significant difference in the efficiency of the extract for antibacterial activity. They showed it as a potent antibacterial agent in the food industry.

Mahmoudi et al. [37] extracted phenolics using methanol from the leaves of different varieties of fig (*Ficus carica* L.). They showed that this methanolic extract had bactericidal activity against Gram-negative and Gram-positive bacteria. They tested *Bacillus cereus* and *Staphylococcus aureus*. They also found that this methanolic extract also exhibited moderate antifungal activity against *Aspergillus brasiliensis* and *Candida albicans*.

Conclusion

Secondary metabolites are important defense tools against many pathogenic microbes. Plants being enriched in these secondary metabolites have high possibility to be used for cure against pathogenic microbes. These may be exploited using biotechnological tools.

Acknowledgements

The facilities of the Department of Biotechnology, Ministry of Science and Technology, Government of India, New Delhi (DBT) under the Bioinformatics Sub Centre and M.Sc. Biotechnology program used in the present work are gratefully acknowledged.

References

1. Falkowska A, Gutowska I, Goschorska M, Nowacki P, Chlubek D, et al. (2015) Energy metabolism of the brain, including the cooperation between astrocytes and neurons, especially in the context of glycogen metabolism. *International J Mol Sci* 16(11): 25959-25981.
2. Haas R, Cucchi D, Smith J, Pucino V, Macdougall CE, et al. (2016) Intermediates of Metabolism: From bystanders to signalling molecules. *Trends Biochem Sci* 41(5): 450-471.
3. Kumar A, Sanwal GG (1988) Kinetics of starch phosphorylase from young banana leaves. *Phytochemistry* 27(4): 983-988.
4. Van der Knaap J, Verrijzer CP (2016) Undercover: gene control by metabolites and metabolic enzymes. *Genes Dev* 30(21): 2345-2369.
5. Freeland-Graves JH, Bavik C (2003) Coenzymes. *Encyclopedia Food Sci Nutr* 1475-1481.
6. Mazid M, Khan TA, Mohammad F (2011) Role of secondary metabolites in defense mechanisms of plants. *Biol Med* 3(2): 232-249.
7. Anulika NP, Ignatius EO, Raymond ES, Osasere OI, Abiola AH (2016) The Chemistry of natural products: plant secondary metabolites. *Intl J Technol Enhancement Emerg Eng Res* 4 (8): 1-8.
8. Kumar V, Shahid M, Srivastava M, Pandey S, Singh A, et al. (2014) Role of secondary metabolites produced by commercial *Trichoderma* species and their effect against soil borne pathogens. *Biosensors J* 3: 1000108. Doi: 10.4172/2090-4967.1000108

9. Selmar SD, Kleinwachter M (2013) Stress enhances the synthesis of secondary plant products: The impact of stress related over reduction on the accumulation of natural products. *Plant Cell Physiol* 54(6): 817-826.
10. Cragg GM Newman DJ (2013) Natural products: A continuing source of novel drug leads. *Biochim. Biophys. Acta* 1830(6): 3670-3695.
11. Delgoda R, Murray JE (2017) Evolutionary perspectives on the role of plant secondary metabolites. In *Pharmacognosy: Fundamentals, Applications and Strategies* 93-100 Academic Press.
12. Praveen M, Ghalib RM (2012) Flavonoids and antimicrobial activity of leaves of *Xylosma longifolium*. *J. Chil. Chem. Soc* 57: 989-991.
13. Tolambiya P, Mathur S (2016) A study on potential phytopharmaceuticals assets in *Catharanthus roseus* L. (Alba). *International J Life Sci Biotechnol Pharma Res* 5: 1-6.
14. Balabirami S and Patharajan S (2012) In vitro antimicrobial and antifungal activity of *Catharanthus roseus* leaves extract against important pathogenic organisms. *International J. Pharmacy Pharmaceut. Sci* 4(3): 487-490.
15. Hassan K, Brenda A, Patrick V, Patrick O (2011) In vivo antidiarrheal activity of the ethanolic leaf extract of *Catharanthus roseus* Linn (Apocyanaceae) in Wistar rats. *African J Pharmacy Pharmacology* 5(5): 1797-1800.
16. Appalasamy S, Lo KY, Ch'ng SJ, Nornadia K, Othman AS, et al. (2014) Antimicrobial activity of artemisinin and precursor derived from *in vitro* plantlets of *Artemisia annua* L. *Biomed. Res. International*. doi: 10.1155/2014/215872
17. IUPAC. Compendium of Chemical Terminology (<http://goldbook.iupac.org/A00220.html>), 2nd ed. (The "Gold Book"). Compiled by A.D. McNaught and A. Wilkinson. Blackwell Scientific Publications, Oxford (1997) ISBN 0-9678550-9-8 doi:10.1351/goldbook
18. Manske RHF (1965) *The Alkaloids. Chemistry and Physiology*. Volume VIII. - New York: Academic Press, p. 673
19. Pavarini DP, Pavarini SP, Niehues M, Lopes NP (2012) Exogenous influences on plant secondary metabolite levels. *Animal Feed Sci Technol* 176(1-4): 5-16.
20. Wink, M. (2003) Alkaloids: Properties and determination. In *Encyclopedia of Food Sciences and Nutrition*. Second edition, Eds. Luiz Trugo and Paul M Finglas. P. 126-134, Elsevier Science Ltd. Publisher.
21. Mojab F (2012) Antimalarial natural products: a review. *Avicenna J. Phytomed* 2(2): 52-62.
22. Chierrito TPC, Aguiar ACC, De Andrade LM, Ceravolo IP, Goncalves RAC, et al. (2014) Anti-malarial activity of indole alkaloids isolated from *Aspidosperma olivaceum*. *Malaria Journal* 13: 142. Doi: 10.1186/1475-2875-13-142
23. Fayed AH (2010) Brain trace element concentration of rats treated with the plant alkaloid, vincamine. *Biol. Trace Elem Res* 136(3): 314-319. Doi: 10.1007/s12011-009-8550-3
24. Tavares LC, Zanon G, Weber AD, Neto AT, Mostardeiro CP, et al. (2014) Structure-activity relationship of benzophenanthridine alkaloids from *Zanthoxylum rhoifolium* having antimicrobial activity. *PLOS One* 9 (5), e97000. Doi: 10.1371/journal.pone.0097000
25. Koul M, Lakra NS, Chandra R, Chandra S (2013) *Cantharanthus roseus* and prospects of its endophytes: a new avenue for production of bioactive metabolites. *Intl J Pharm Sc. Res* 4(7): 2705-2716.
26. Jennifer Mordue A, Nisbet AJ (2000) Azadirachtin from the Neem tree *Azadirachta indica*: its action against insects. *An Soc. Entomol. Brasil* 29: 615-632.
27. Kim WS, Choi WJ, Lee S, Kim WJ, Lee DC, et al. (2015) Anti-inflammatory, Anti-oxidant and Anti-microbial effects of Artemisinin extracts from *Artemisia annua* L. *Korean J Physiol. Pharmacol* 19(1) 21-27. Doi: 10.4196/kipp.2015.19.1.21
28. Mitscher LA, Park YH, Clark D, Beal JL (1980) Antimicrobial agents from higher plants. Antimicrobial isoflavonoids and related substances from *Glycyrrhiza glabra* L. var. *typica*. *J. Nat. Prod* 43(3): 259-269.
29. Dastidar SG, Manna A, Kumar KA, Mazumdar K, Dutta NK, et al. (2004) Studies on the antibacterial potentiality of isoflavones. *International J Antimicrob Agent* 23(1): 99-102.
30. Mukne AP, Viswanathan V, Phadatare AG (2011) Structure pre-requisites for isoflavones as effective antibacterial agents. *Pharmacogn Rev* 5(9): 13-18.
31. Praveen M, Ghalib RM (2012) Flavonoids and antimicrobial activity of leaves of *Xylosma longifolium*. *J. Chil. Chem. Soc.* 57 : 989-991.
32. Maddox CE, Laur LM, Tian L (2010) Antibacterial activity of phenolic compounds against the phytopathogen *Xyella fastidiosa*. *Curr. Microbiol* 60(1): 53-58.
33. Papuc C, Goran GV, Predescu CN, Nicorescu V, Stefan G (2017) Plant polyphenols as antioxidant and antibacterial agents for shelf life extension of meat and meat products: Classification, structure, sources and action mechanisms *Compreh. Rev Food Sci Food Safety* 16: 1243-1268.

-
-
34. Gulluce M, Sokmen M, Daferera D, Agar G, Ozkan H, et al. (2003) In vitro antibacterial, antifungal and antioxidant activities of the essential oil and methanol extracts of herbal parts and callus cultures of *Saturja hortensis* L. *J. Agric. Food Chem.* 51(2): 3958-3965.
 35. Cui Y, Oh YJ, Lim J, Youn M, Lee I, et al. (2012) AFM study of the differential inhibitory effects of the green tea polyphenol (-)-epigallocatechin-3-gallate (EGCG) against Gram-positive and Gram-negative bacteria. *Food Microbiol.* 29(1): 80-87.
 36. Ouerghemmi I, Rebey IB, Rohali FZ, Bourgou S, Pistelli L, et al. (2017) Antioxidant and antimicrobial phenolic compounds from extracts of cultivated and wild grown Tunisian *Ruta chalepensis*. *J. Food Drug Analysis* 25(2) : 350-359.
 37. Mahmoudi S, Khali M, Benkhaled A, Benamirouche K, Baiti I (2016) Phenolic and flavonoid contents, antioxidant and antimicrobial activities of leaf extracts from ten Algerian *Ficus carica* L. varieties. *Asian Pac J Trop Biomed* 6(3): 239-245.